International Food Research Journal 29(1): 142 - 148 (February 2022)

Journal homepage: http://www.ifrj.upm.edu.my



Improving oxidative stress and inflammation status of obese women with metabolic syndrome using phenolic-rich red kidney bean sprout milk yogurt

¹*Winarsi, H. and ²Septiana, A. T.

¹Department of Nutrition Science, Faculty of Health Sciences, Universitas Jenderal Soedirman, Purwokerto, 53123, Central Java, Indonesia ²Department of Food Technology, Faculty of Agriculture, Universitas Jenderal Soedirman, Purwokerto, 53123, Central Java, Indonesia

Article history

Received: 6 April 2020 Received in revised form: 29 March 2021 Accepted: 9 July 2021

Keywords

SOD, TNF-a, BMI, phenolic, red kidney bean sprout milk yogurt

Abstract

Oxidative stress and inflammation are involved in the pathogenesis of metabolic syndrome (MetS). Antioxidant-rich food products are known for improving the oxidative stress and inflammation as well as inhibiting the development of metabolic syndrome. The present work thus aimed to determine the effects of phenolic-rich red kidney bean sprout milk yogurt (RKBSMY) on superoxide dismutase (SOD) activity, TNF- α level, and body mass index (BMI) of women with MetS. Thirty obese women with MetS, low SOD activity, and high TNF- α level served as research subjects. The 2-month intervention using red kidney bean sprout milk yogurt (RKBSMY) with 2% lactic acid bacterial starter, 10% sucrose, and 24-h fermentation was conducted to women with MetS. Results showed that RKBSMY increased the SOD activity from 5.13 to 8.02 ng/mL (p = 0.047), decreased the plasma TNF- α level from 60.89 to 39.77 pg/mL (p < 0.05), and decreased the BMI from 28.04 to 24.38 kg/m². Therefore RKBSMY could be beneficial for people with degenerative diseases associated with oxidative stress

© All Rights Reserved

Introduction

Various factors can trigger the emergence of metabolic syndrome (MetS) which is a group of simultaneously occurring metabolic disorders. These disorders include the increase in high blood pressure, accumulation of fat in stomach, increase in blood sugar, and increase in cholesterol and triglyceride levels. These are usually triggered by the oxidative stress and inflammatory conditions.

The development of MetS may also lead to type 2 diabetes mellitus and cardiovascular disease (Alshehri, 2010). Montague and O'Rahilly (2000) added that obesity is a major MetS-causing component; however, its role is still unclear. Under the obesity conditions, adipose cells produce TNF-α active molecule, and release both interleukin-6 (IL-6) and C-reactive protein (CRP), as well as pro-inflammatory cytokines (Winarsi et al., 2016). TNF-α has been recognised as a pro-inflammatory cytokine which has an important role in the pathogenesis of a number of chronic inflammatory diseases. The increase in TNF- α contributes to the development of thrombosis and insulin resistance (Nair et al., 2006). Therefore, it is important to control inflammation since it is believed as the key to the development of MetS.

Additionally, obesity also triggers the systemic oxidative stress associated with high body mass index (BMI) and accumulated fat which further underlies the dysregulation of adipocytokine and development of MetS. Winarsi et al. (2012) reported that high oxidative stress is mostly found in women aged > 40 years. This population subgroup often has low activity of superoxide dismutase (SOD) and high levels of CRP and TNF-α (Winarsi et al., 2015). Shahbazian et al. (2013) said that the risk of MetS in women is higher than in men. Some researchers reported that oxidative stress interferes with glucose absorption (Krishna et al., 2015), decreases insulin secretion, underlies hypertension, and even damages blood vessel walls (Wu et al., 2014). It is therefore imperative to tackle oxidative stress and reduce BMI of people with MetS.

Food products rich in phenolic antioxidants can improve oxidative stress, including increasing the activity of antioxidant enzymes such as SOD (Winarsi *et al.*, 2016; Şenay *et al.*, 2019), inflammation status (including decreasing the levels of cytokines such as IL-6, CRP, and TNF-alpha) (Yehuda *et al.*, 2015), and normalisation of women's weight (Guo *et al.*, 2017). Winarsi *et al.* (2019)

reported that red kidney bean sprout milk yogurt (RKBSMY) contained 993.08 ppm of phenolic antioxidants.

Phenolic acts as the direct scavengers of superoxide anion by cutting the elongation of free-radical oxidation chain or scavenging the free radicals (Rosignoli et al., 2013). Flavonoids, one of the phenolic compounds, have antiradical properties especially against hydroxyl, superoxide anions, peroxyl, and alkoxyl radicals. The presence of these flavonoid compounds has a very strong affinity for Fe ions, a catalyst for free-radical formation (Fu et al., 2013). Phenolics also have anti-inflammatory properties by inhibiting the synthesis of pro-inflammatory mediators, modifying eicosanoid, inhibiting the activation of immune cells, and inhibiting the activity of nitric oxide synthase (NOS) and cyclooxygenase-2 (Chuang and McIntosh, 2011). The research question of the present work was whether or not the phenolic content of RKBSMY could improve the antioxidant status, inflammation status, and BMI in women with MetS. The present work thus aimed to determine the effects of RKBSMY on SOD activity, TNF-α level, and BMI of women with MetS.

Materials and methods

The present work employed a true experimental design, and has obtained the ethical approval from the Health Research Ethics Commission (*Komisi Etik Penelitian Kesehatan; KEPK*) from the Faculty of Medicine, Jenderal Soedirman University (reference no.: 3343/KEP-K/VII/2019).

Production of RKBSMY

Sucrose (10%) and skim milk (10%) were added to red kidney bean sprout milk (RKBSM). The milk was pasteurised at 70°C for 10 min, and then cooled to 45°C. The milk was inoculated with commercial lactic acid bacteria (LAB) starters at 2% of milk's total volume. The milk was then incubated at the room temperature (27 - 35°C) for 24 h (Winarsi *et al.*, 2019).

Research subjects

A total of 30 women with MetS served as the research subjects with the following inclusion criteria: aged 45 - 80 years, having central obesity, inflammation, and oxidative stress, not leaving the research site for two months, and willing to sign the consent form. The subjects were randomly divided into three groups with a total of 10 people in each

group.

RKBSMY intervention in research subjects

The first group (control) only took medication from doctor; the second group received placebo treatment; and the third group received 200 mL/d of RKBSMY regularly for two months.

Blood sampling

Blood samples (1 mL) of research subjects were taken three times (0, 1, and 2 months after the intervention). The blood samples were centrifuged at 3,000 rpm for 10 min. The blood plasma was then examined to determine the SOD activity (using Fine Test Human SOD ELISA Kits) and TNF- α level (using Fine Test Human TNF-alpha ELISA Kits). Weight (kg) and height (m) measurements were also conducted every time the blood samples were taken. BMI was calculated by dividing the weight (kg) by the height square (m²).

Data analysis

The obtained data were analysed using *F*-test, and continued with Duncan's Multiple Range Test (DMRT) at the level of 5% if there was a significant difference.

Results and discussion

Effects of RKBSMY on SOD activity

RKBSMY increased SOD activity from 5.13 to 8.02 ng/mL (p = 0.047). The longer the intervention duration, the higher the SOD activity. The highest activity occurred after 2-months intervention in the treatment group, but SOD activity decreased in the control (p = 0.0036) and placebo group (p = 0.0058) (Figure 1).

These results are similar to the findings of Kojadinovic *et al.* (2016), who also added that 300 mL/d of pomegranate juice given for six weeks could reduce the thiobarbituric acid reactive substances in erythrocytes. A 5-year Mediterranean diet with olive oil or legume extracts also increased the plasma SOD and catalase activity (Sureda *et al.*, 2016). The increase in SOD activity is possibly related to the phenolic antioxidant content in RKBSMY.

The antioxidant potential of phenolic compounds is determined by the chemical structure, number, and position of hydroxyl groups, conjugated groups, degree of glycosylation, and presence of electron donors in the ring structure (Kumar and Pandey, 2013). It is said that phenolic bioavailability is low, but its metabolism and elimination are high.

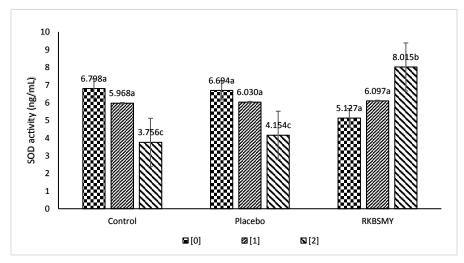


Figure 1. Effect of red kidney bean sprout milk yogurt (RKBSMY) on plasma SOD activity in women with metabolic syndrome (MetS). [0] = baseline data (before receiving the intervention); [1] = data obtained after subjects received the intervention for one month; and [2] = data obtained after subjects received the intervention for two months.

Therefore, its antioxidant effects potentially have clinical relevance. Polyphenols, one of which is phenolic, are an important dietary antioxidant. They are biomolecules found in plant products, and proven able to exert antioxidant and anti-inflammatory effects both in vitro and in vivo (Chiva-Blanch and Visioli, 2012). According to Ovaskainen et al. (2008), the total phenolic intake is generally estimated at 900 mg/d. In the present work, the subjects received RKBSMY at 200 mL/d, which is equivalent to 198.8 mg of phenolic/d (Winarsi et al., 2019), and only 22% from the opinion of Ovaskainen et al. (2008), but proven able to increase SOD activity to 56.33%. In other words, phenolic RKBSMY is a potential antioxidant because it can induce the performance of plasma SOD enzyme in MetS sufferers.

There are several mechanisms by which polyphenols could increase SOD activity. Hu *et al.* (2019) have reported that polyphenols increase the expression of stress-response proteins (*e.g.*, heat shock proteins) and SOD antioxidant enzymes, and thereby suppress the reactive oxygen species (ROS) level. Polyphenols minimise the oxidative stress by cleaning the ROS, and producing more stable phenolic radicals. The radical scavenging ability of polyphenols mainly comes from the D ring in the galloyl structure (Severino *et al.*, 2009). Hydroxyl radicals and ROS are bound by polyphenols, especially through the oxidised D-ring galloyl group.

In addition, polyphenols also have indirect antioxidant effects. As reported by Feng *et al.* (2002), the administration of tea polyphenols could reduce the 8-oxoguanine level (a marker for DNA damage caused by oxidative stress), and inhibit

DNA oxidative damage (by reducing the expression of cytochrome P450). Therefore, polyphenols could mitigate the symptoms related to oxidative stress by increasing antioxidant capacity.

Phenolic prevents oxidation by chelating the metal ions, and inhibiting the reaction of Fenton and Haber-Weis as the sources of oxygen free radicals. Phenolics can directly scavenge superoxide radicals (Winarsi et al., 2016). It is possible that phenolic in RKBSMY could have contributed to reducing the superoxide radical reactivity at early stage, so that the compound could not turn into hydrogen peroxide which has the potential to become a radical. Therefore, there could be no oxidant build-up as indicated by the increased activity of SOD. Furthermore, the subjects received 200 mL/d of RKBSMY, which was equivalent to 198.9 mg of phenolic daily (Winarsi et al., 2019). The plasma SOD activity in women with MetS increased by 56.33% at this dosage.

The effect of RKBSMY on TNF-α level

After 2-month intervention using RKBSMY, the plasma TNF- α level in women with MetS significantly decreased (p < 0.05) from 60.89 to 39.77 pg/mL (Figure 2). Besides its effect as an antioxidant that suppresses the oxidative stress, it turns out that the phenolic in RKBSMY also showed anti-inflammatory effects. The phenolic compounds affect the inflammation-related cell signalling pathways and gene expression. Phenolics are mostly found in beans and their processed products, including the yogurt made from red kidney bean sprouts used in the present work.

The half-life of polyphenol elimination is

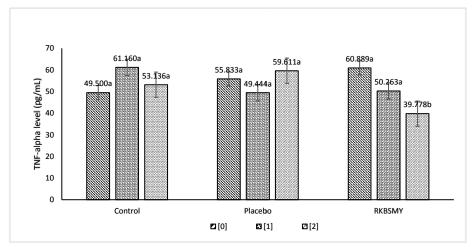


Figure 2. Effect of red kidney bean sprout milk yogurt (RKBSMY) on plasma TNF-alpha levels in women with metabolic syndrome (MetS). [0] = baseline data (before receiving the intervention); [1] = data obtained after subjects received the intervention for one month; and [2] = data obtained after subjects received the intervention for two months.

between 1 and 18 h, but most polyphenols can be excreted in less than 8 h (Manach *et al.*, 2005). This finding proves that most polyphenols are excreted from the body within one day after consumption. Therefore, the concentration of plasma and tissue polyphenols depends strongly on the consumption patterns, and their levels in plasma also vary throughout the day. The 2-month intervention utilised in the present work allowed the phenolic in RKBSMY to affect the inhibition of inflammation development, as indicated by the decrease in TNF- α level.

Some studies have shown that flavonoids, one of the phenolic types, modulate several inflammatory response mediators. Flavonoids inhibit the expression of pro-inflammatory genes (e.g., cytokines, chemokines, or adhesion molecules) and that of inducible NOS (iNOS) isoforms (García-Lafuente et~al., 2009). The administration of products containing quercetin, one of the phenolic compounds, reduced the TNF- α in obese patients (Egert et~al., 2009).

González *et al.* (2011) confirmed that the anti-inflammatory activity of flavonoids occurs through several mechanisms; *i.e.*, inhibition of cyclooxygenase (COX) and lipoxygenase activities, inhibition of leukocyte accumulation, inhibition of neutrophil degranulation, and inhibition of histamine release. Inflammation occurs due to the release of chemical mediators from the damaged tissue and cell migration. Polyphenols are known able to inhibit the inflammation associated with cytokine production. According to Nair *et al.* (2006), polyphenols are one of natural ingredients which could inhibit the activation of NF-kB, thus decreasing TNF-α production. It has also been stated that polyphenol

activity is specific, depending on the type of cell and its structure. Polyphenols block the release of TNF- α by modulating the MAPK pathway at various levels of signalling pathways. In epithelial cells, polyphenols block the TNF- α , and trigger the ICAM-1 by inhibiting the ERK, JNK, and p38 (Nizamutdinova *et al.*, 2007). The ability of polyphenol compounds to block the MAPK pathway makes this bioactive substance potentially therapeutic, which protects the body from the inflammatory reactions.

Concerning its role in the decrease of inflammatory cytokines, cyclooxygenase and lipoxygenase are two enzymes which have an important role as the inflammatory mediators. Both enzymes are released by the oxidised arachidonic acid, together with the initial inflammatory response. Neutrophils contain lipoxygenase which produces chemotactic compounds from the arachidonic acid to release cytokines (Kojadinovic *et al.*, 2016). Phenolic compounds can inhibit both cyclooxygenase and lipoxygenase pathways, thereby suppressing the formation of inflammatory metabolites (TNF- α).

The effect of RKBSMY on BMI

Subjects had an average BMI of 28.04 kg/m² at the beginning of the study, thus categorised as obese. The higher the BMI, the wider the surface of adipose tissue. This triggers the O₂ deficiency. Adipose is encouraged to release the pro-inflammatory cytokines, chemokines, and angiogenic factors under this condition to increase blood flow and vascularity (Donath and Shoelson, 2011). It is obvious that high BMI is closely related to inflammatory conditions. Prolonged (chronical) hypoxia causes a chronic release of pro-inflammatory

cytokines. Therefore, it is important to lower or normalise the BMI.

In the present work, RKBSMY decreased the BMI from 28.04 to 24.38 kg/m2. The decrease (11.57%) was greater than in the placebo (2.91%) and control (2.20%) groups. Although the decrease in BMI caused by the RKBSMY was not significant, the decreased BMI now placed the subjects in the normal weight category. Rupasinghe et al. (2016) stated that phenolic compounds can change fat and energy metabolisms, thereby facilitating weight loss and preventing weight gain. The decreasing BMI in women with MetS was also associated with the decreasing level of pro-inflammatory cytokine (TNF-α). Flavonoids act in the management of obesity by controlling hunger and satiety. Kim et al. (2013) confirmed that phenolic compounds, especially in the form of aglycones, could increase the secretion of cholecystokinin (CCK), a hormone secreted by endocrine cells in small intestine. Increasing CCK helps to control the amount of food intake. CCK encourages the release of enzymes in pancreas and bile in the gall bladder. Therefore, CCK also has the function to delay hunger and prolong satiety.

Dallas *et al.* (2008) reported that 20 mg/mL of phenolic supplement could stimulate lipolysis in adipocytes, which were then verified as free fatty acids. Phenol also has an inhibitory effect on cAMP-phosphodiesterase (Dell'Agli *et al.*, 2008). Phosphodiesterase is an enzyme that can convert cAMP to 5'AMP. The amount of cAMP increases with the inhibition of cAMP-phosphodiesterase by phenolic. High cAMP stimulates the action of protein kinase which in turn activates lipase, thus resulting in lipolysis and a decrease in fat content. The low-fat content could also lead to weight loss.

There are several mechanisms of weight loss by polyphenols. One of the mechanisms is their action in gastrointestinal tract by suppressing the digestion and absorption of macronutrients or changing the intestinal microbiota. Polyphenols inhibit anabolism after the systemic absorption occurs with the changes of intestinal microbiota. Polyphenols then stimulate catabolism in liver, muscles, adipose tissue, and other tissues (Yang *et al.*, 2015). Through these mechanisms, polyphenols can decrease body weight, improve MetS condition, and eventually reduce the risk of MetS development.

Regarding the consumption of polyphenol-rich products and their benefits in weight loss, some researchers reported that these antioxidant compounds increase lipid and total nitrogen levels in faeces, which means that

polyphenols can reduce the digestion and absorption of lipid and protein (Yang and Hong, 2013). After the phenolic-rich product is consumed, lipids are then emulsified, hydrolysed, and absorbed in small intestine. Lipid transporters on the apical surface of intestine facilitate the transfer of fatty acids and cholesterol into enterocytes. The absorbed lipids are then packaged into the chylomicrons, and secreted into the lymphatic system. In this case, polyphenols interfere with the emulsification, and inhibit the action of pancreatic lipase and phospholipase, thus reducing the weight gain in rats fed with a high-fat diet (Grove et al., 2012). The mechanism of polyphenols in inhibiting the protein digestion and absorption has not been frequently studied. However, there is a strong relationship between antioxidant, digested protein, and proteinase in the digestive system through H-bonds and hydrophobic interactions, thus resulting in the obstruction of the digestive working system.

The polyphenol inhibitory activity on digestive enzymes occurs in α-amylase, glucosidase, and glucose transporters. Polyphenols inhibit the glucose absorption in the intestinal cells through the competitive inhibitors of sodium-glucose co-transporter 1 (Park et al., 2009). It has also been reported that consumption of polyphenol-rich products can reduce the glucose digestion and absorption due to the inhibition of α-amylase activity by these compounds (Forester et al., 2012). The nutrient-decreasing digestion and absorption have an important role in weight loss, which ultimately inhibit the development of MetS.

Conclusion

Phenolic-rich RKBSMY increased SOD activity by 56.33%, decreased plasma TNF- α level by 34.67%, and decreased BMI of obese women with MetS by 11.57%, which led to normal weight. Therefore, RKBSMY could be beneficial as a healthy and functional food, and source of antioxidants and anti-inflammatory for obese women with MetS.

Acknowledgement

The present work was financially supported by the Directorate General of Development and Research Enhancement of the Ministry of Research, Technology, and Higher Education of Indonesia through the Competency-based Research Scheme (grant no.: P/1782/UN23/14PN/2019 on May 2, 2019).

References

- Alshehri, A. M. 2010. Metabolic syndrome and cardiovascular risk. Journal of Family and Community Medicine 17(2): 73-78.
- Chiva-Blanch, G. and Visioli, F. 2012. Polyphenols and health: moving beyond antioxidants. Journal of Berry Research 2: 63-71.
- Chuang, C. C. and McIntosh, M. K. 2011. Potential mechanisms by which polyphenol-rich grapes prevent obesity-mediated inflammation and metabolic diseases. Annual Review of Nutrition 31: 155-176.
- Dallas, C., Gerbi, A., Tenca, G., Juchaux, F. and Bernard, F.-X. 2008. Lipolytic effect of a polyphenolic citrus dry extract of red orange, grapefruit, orange (SINETROL) in human body fat adipocytes. Mechanism of action by inhibition of cAMP-phosphodiesterase (PDE). Phytomedicine 15(10): 783-792.
- Dell'Agli, M., Maschi, O., Galli, G. V., Fagnani, R.,
 Dal Cero, E., Caruso, D. and Bosisio, E. 2008.
 Inhibition of platelet aggregation by olive oil phenols via cAMP-phosphodiesterase. British Journal of Nutrition 99(5): 945-951.
- Donath, M. Y. and Shoelson, S. E. 2011. Type 2 diabetes as an inflammatory disease. Nature Reviews Immunology 11(2): 98-107.
- Egert, S., Boesch-Saadatmandi, C., Wolffram, S., Rimbach, G. and Müller, M. J. 2009. Serum lipid and blood pressure responses to quercetin vary in overweight patients by apolipoprotein E genotype. Journal of Nutrition 140(2): 278-284.
- Feng, Q., Torii, Y., Uchida, K., Nakamura, Y., Hara, Y. and Osawa, T. 2002. Black tea polyphenols, theaflavins, prevent cellular DNA damage by inhibiting oxidative stress and suppressing cytochrome P450 1A1 in cell cultures. Journal of Agricultural and Food Chemistry 50(1): 213-220.
- Forester, S. C., Gu, Y. and Lambert, J. D. 2012. Inhibition of starch digestion by the green tea polyphenol, (-)-epigallocatechin-3-gallate. Molecular Nutrition and Food Research 56(11): 1647-1654.
- Fu, Y., Chen, J., Li, Y.-J., Zheng, Y.-F. and Li, P. 2013. Antioxidant and anti-inflammatory activities of six flavonoids separated from licorice. Food Chemistry 141(2): 1063-1071.
- García-Lafuente, A., Guillamón, E., Villares, A., Rostagno, M. A. and Martínez, J. A. 2009. Flavonoids as anti-inflammatory agents: implications in cancer and cardiovascular disease. Inflammation Research 58: 537-552.

- González, R., Ballester, I., López-Posadas, R., Suárez, M. D., Zarzuelo, A., Martínez-Augustin, O. and Medina, F. S. D. 2011. Effects of flavonoids and other polyphenols on inflammation. Critical Reviews in Food Science and Nutrition 51(4): 331-362.
- Grove, K. A., Sae-Tan, S., Kennett, M. J. and Lambert, J. D. 2012. (-)-Epigallocate-chin-3-gallate inhibits pancreatic lipase and reduces body weight gain in high fat-fed obese mice. Obesity 20(11): 2311-2313.
- Guo, X., Tresserra-Rimbau, A., Estruch, R., Martínez-González, M. A., Medina-Remón, A., Fitó, M., ... and Lamuela-Raventós, R. M. 2017. Polyphenol levels are inversely correlated with body weight and obesity in an elderly population after 5 years of follow up (the randomised PREDIMED study). Nutrients 9(5): article no. 452.
- Hu, R., He, Y., Arowolo, M., Wu, S. and He, J. 2019. Polyphenols as potential attenuators of heat stress in poultry production. Antioxidants 8(3): article no. 67.
- Kim, H. Y., Park, M., Kim, K., Lee, Y. M. and Rhyu,
 M. R. 2013. Hesperetin stimulates cholecysto-kinin secretion in enteroendocrine STC-1 cells. Biomolecules and Therapeutics 21(2): 121-125.
- Kojadinovic, M. I., Arsic, A. C., Debeljak-Martacic, J. D., Konic-Ristic, A. I., Kardum, N. D., Popovic, T. B. and Glibetic, M. D. 2016. Consumption of pomegranate juice decreases blood lipid peroxidation and levels of arachidonic acid in women with metabolic syndrome. Journal of the Science of Food and Agriculture 97(6): 1798-1804.
- Krishna, M. S., Joy, B. and Sundaresan, A. 2015. Effect on oxidative stress, glucose uptake level and lipid droplet content by Apigenin 7, 4'-dimethyl ether isolated from *Piper longum* L. Journal of Food Science and Technology 52(6): 3561-3570.
- Kumar, S. and Pandey, A. K. 2013. Chemistry and biological activities of flavonoids: an overview. The Scientific World Journal: article ID 162750
- Manach, C., Williamson, G., Morand, C., Scalbert, A. and Rémésy, C. 2005. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. American Journal of Clinical Nutrition 81(Suppl 1): 230S-242S.
- Montague, C. T. and O'Rahilly, S. 2000. The perils of portliness: causes and consequences of visceral adiposity. Diabetes 49: 883-888.
- Nair, M. P., Mahajan, S., Reynolds, J. L., Aalinkeel,

- R., Nair, H., Schwartz, S. A. and Kandaswami, C. 2006. The flavonoid quercetin inhibits proinflammatory cytokine (tumor necrosis factor alpha) gene expression in normal peripheral blood mononuclear cells via modulation of the NF- $\kappa\beta$ system. Clinical and Vaccine Immunology 13(3): 319-328.
- Nizamutdinova, I. T., Oh, H. M., Min, Y. N., Park, S. H., Lee, M. J., Kim, J. S., ... and Kim, H. J. 2007. Paeonol suppresses intercellular adhesion molecule-1 expression in tumor necrosis factor-α-stimulated human umbilical vein endothelial cells by blocking p38, ERK and nuclear factor-κB signaling pathways. International Immunopharmacology 7(3): 343-350.
- Ovaskainen, M. L., Torronen, R., Koponen, J. M., Sinkko, H., Hellstrom, J., Reinivuo, H. and Mattila, P. 2008. Dietary intake and major food sources of polyphenols in Finnish adults. Journal of Nutrition 138: 562-566.
- Park, J. H., Jin, J. Y., Baek, W. K., Park, S. H., Sung, H. Y., Kim, Y. K., ... and Song, D. K. 2009.
 Ambivalent role of gallated catechins in glucose tolerance in humans: a novel insight into non-absorbable gallated catechin-derived inhibitors of glucose absorption. Journal of Physiology and Pharmacology 60(4): 101-109.
- Rosignoli, P., Fuccelli, R., Fabiani, R., Servili, M. and Morozzi, G. 2013. Effect of olive oil phenols on the production of inflammatory mediators in freshly isolated human monocytes. The Journal of Nutritional Biochemistry 24(8): 1513-1519.
- Rupasinghe, H. P. V., Sekhon-Loodu, S., Mantso, T. and Panayiotidis, M. I. 2016. Phytochemicals in regulating fatty acid β -oxidation: potential underlying mechanisms and their involvement in obesity and weight loss. Pharmacology and Therapeutics 165: 153-163.
- Şenay, S., Islim, P. and Tugay, A. 2019. Supplementation of natural antioxidants to reduced crude protein diets for Japanese quails exposed to heat stress. Brazilian Journal of Poultry Science 21(1): article no. 694.
- Severino, J. F., Goodman, B. A., Kay, C. W. M., Stolze, K., Tunega, D., Reichenauer, T. G. and Pirker, K. F. 2009. Free radicals generated during oxidation of green tea polyphenols: electron paramagnetic resonance spectroscopy combined with density functional theory calculations. Free Radical Biology and Medicine 46(8): 1076-1088.
- Shahbazian, H., Latifi, S. M., Jalali, M. T., Shahbazian, H., Amani, R., Nikhoo, A. and

- Aleali, A. M. 2013. Metabolic syndrome and its correlated factors in an urban population in South West of Iran. Journal of Diabetes and Metabolic Disorders 12(1): article no. 11.
- Sureda, A., del Mar Bibiloni, M., Martorell, M., Buil-Cosiales, P., Marti, A., Pons, A., ... and Gonzalez, M. A. M. 2016. Mediterranean diets supplemented with virgin olive oil and nuts enhance plasmatic antioxidant capabilities and decrease xanthine oxidase activity in people with metabolic syndrome: the PREDIMED study. Molecular Nutrition and Food Research 60(12): 2654-2664.
- Winarsi, H., Sasongko, N. D. and Purwanto, A. 2016. Germinated-soy milk in supressing inflammation and oxidative stress in blood plasma and breast milk of lactating mothers. International Food Research Journal 23(2): 646-652.
- Winarsi, H., Septiana, A. T., Kartini and Hanifah, I. N. 2019. Fermented lactic acid bacteria increases the phenolic content and fiber of red kidney bean sprouts (*Phaseolus vulgaris* L) yogurt, a functional drink for obesity. Jurnal Gizi dan Pangan Soedirman 3(1): 64-75.
- Winarsi, H., Wijayanti, S. P. M. and Purwanto, A. 2012. Soy germ protein with or without-Zn improve plasma lipid profile in metabolic syndrome women. HAYATI Journal of Biosciences 19(1): 25-30.
- Winarsi, H., Wijayanti, S. P. M., Sasongko, N. D. and Purwanto, A. 2015. Effects of soy-germ protein on catalase activity of plasma and erythrocyte of metabolic syndrome women. HAYATI Journal of Biosciences 22(1): 1-5.
- Wu, J., Xia, S., Kalionis, B., Wan, W. and Sun, T. 2014. The role of oxidative stress and inflammation in cardiovascular aging. BioMed Research International 2014: article ID 615312.
- Yang, C. S. and Hong, J. 2013. Prevention of chronic diseases by tea: possible mechanisms and human relevance. Annual Review of Nutrition 33(1): 161-181.
- Yang, C. S., Zhang, J., Zhang, L., Huang, J. and Wang, Y. 2015. Mechanisms of body weight reduction and metabolic syndrome alleviation by tea. Molecular Nutrition and Food Research 60(1): 160-174.
- Yehuda, I., Madar, Z., Leikin-Frenkel, A. and Tamir, S. 2015. Glabridin, an isoflavan from licorice root, downregulates iNOS expression and activity under high-glucose stress and inflammation. Molecular Nutrition and Food Research 59: 1041-1052.